




REVIEW

Controversies in the pathophysiology of leg ulcers in sickle cell disease

Judith Catella^{1,2,3,4}   | Nicolas Guillot^{2,3} | Elie Nader^{2,3} | Sarah Skinner⁵ | Solène Poutrel^{1,2,3} | Arnaud Hot^{1,2,3} | Philippe Connes^{2,3}  | Berengère Fromy⁴

¹Service de Médecine Interne et Vasculaire, Hôpital Edouard Herriot, Hospices Civils de Lyon, Lyon, France

²Laboratoire d'Excellence du Globule Rouge (Labex GR-Ex), Sorbonne, Paris, France

³Laboratoire Interuniversitaire de Biologie de la Motricité (LIBM) EA7424, Equipe "Biologie Vasculaire et du Globule Rouge", Université Claude Bernard Lyon 1, Université de Lyon, Lyon, France

⁴Laboratoire de Biologie Tissulaire et Ingénierie Thérapeutique (LBTI UMR 5305), CNRS/Université Claude Bernard Lyon 1, Lyon, France

⁵Clinical Research and Epidemiology Unit, Montpellier University, Montpellier, France

Correspondence

Judith Catella, Service de Médecine Interne et Médecine Vasculaire, Hôpital Edouard Herriot, Hospices Civils de Lyon, 69003 Lyon, France.

Email: judith.catella@chu-lyon.fr

Summary

Patients with sickle cell disease (SCD) often experience painful vaso-occlusive crises and chronic haemolytic anaemia, as well as various acute and chronic complications, such as leg ulcers. Leg ulcers are characterized by their unpredictability, debilitating pain and prolonged healing process. The pathophysiology of SCD leg ulcers is not well defined. Known risk factors include male gender, poor social conditions, malnutrition and a lack of compression therapy when oedema occurs. Leg ulcers typically start with spontaneous pain, followed by induration, hyperpigmentation, blister formation and destruction of the epidermis. SCD is characterized by chronic haemolysis, increased oxidative stress and decreased nitric oxide bioavailability, which promote ischaemia and inflammation and consequently impair vascular function in the skin. This cutaneous vasculopathy, coupled with venostasis around the ankle, creates an ideal environment for local vaso-occlusive crises, which can result in the development of leg ulcers that resemble arterial ulcers. Following the development of the ulcer, healing is hindered as a result of factors commonly observed in venous ulceration, including venous insufficiency, oedema and impaired angiogenesis. All of these factors are modulated by genetic factors. However, our current understanding of these genetic factors remains limited and does not yet enable us to accurately predict ulceration susceptibility.

KEYWORDS

chronic wounds, leg ulcers, pathophysiology, sickle cell disease, treatment

INTRODUCTION

Sickle cell disease (SCD) is an autosomal recessive genetic disorder secondary to a single mutation in the β -globin gene. Sickle cell anaemia (SCA; HbSS) is the most common and the most severe form of SCD.¹ Data from the United States, from 1989 to 2012, report that, for every 1941 newborn births screened, one case of SCD was identified and one of every 67 neonates screened was heterozygous for the beta-S allele.²

The primary mechanism at the origin of the complex SCD pathophysiology is HbS polymerization in deoxygenated conditions,³ which causes the sickling of red blood cells

(RBCs). Sickled RBCs are poorly deformable and very fragile, putting patients with SCD at risk for repeated painful vaso-occlusive crises and chronic haemolytic anaemia.⁴ Patients with SCD are not only at risk of developing various acute complications but may also develop chronic complications, including leg ulcers. Chronic leg ulceration in SCD has been associated with significant clinical and psychosocial morbidity⁵⁻⁷ and an increased risk of early death.⁸ Prevalence of chronic leg ulcers among individuals with SCD varies from 1.4% in Brazil to 29.5% in Jamaica.⁹⁻¹³

In order to improve the management and care of SCD leg ulcers, the mechanisms behind their occurrence must be

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial-NoDerivs](https://creativecommons.org/licenses/by-nc-nd/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2024 The Author(s). *British Journal of Haematology* published by British Society for Haematology and John Wiley & Sons Ltd.



better understood. In this review, we aim to explore several contentious aspects of the current understanding of leg ulcer pathophysiology in SCD and examine potential factors contributing to their delayed healing.

SCD ULCERS START MOSTLY LIKE AN ISCHAEMIC ULCER

Ulcerations in SCD are likely caused by local vaso-occlusive crises, resulting in cutaneous microvascular infarctions. The majority of SCD ulcerations manifest spontaneously, without any associated trauma.⁵ Patients usually experience the initial symptom of spontaneous pain, followed by induration, hyperpigmentation or subcutaneous necrosis, blister formation and destruction of the epidermis.⁵ In some patients, leg ulcers may occur after minor trauma or scratching.^{5,14} However, irrespective of the mode of onset, the progression of the ulcer tends to follow a similar course.⁵

General conditions

Several studies reported a high incidence of leg ulcers in males compared to females with SCD.^{11,13,15-18} The male to female ratio was approximately 2 in the United States and Nigeria.^{16,17} In contrast, other studies reported a similar incidence between males and females.^{9,12,15,19} Thus, no clear conclusion about the effect of gender on ulceration can be drawn from the literature.

In addition, it has been observed that patients with leg ulcers tend to have poorer social conditions.¹⁰ Cumming et al. reported that, in a Jamaican cohort of individuals with SCD, 85% of patients with leg ulcers were below the poverty line, whereas only 46% of those without leg ulcers fell below the poverty line.⁹ It remains challenging to determine whether the incidence of leg ulcers is related to housing or working conditions. As discussed below, oedema can delay healing,²⁰ and factors such as prolonged standing or sleeping in a seated position can exacerbate oedema.²¹ SCD patients with leg ulcers should be questioned about their living conditions (Figure 1). Physicians should emphasize the importance of leg care to their patients and specify that if oedema occurs, they should elevate their legs whenever possible to prevent leg ulcers and minimize healing delays. Depending on the country in question, social workers may have varying capacities to assist SCD patients with housing conditions and financial support.

In SCD patients, poor social conditions may also contribute to malnutrition. Malnutrition, which can be characterized by a low serum albumin level¹⁹ or a low body mass index (BMI),^{11,19} has been linked to the occurrence of leg ulcers in SCD patients. In general, malnutrition, a common factor among patients suffering from wounds,²² may precede the appearance of ulcers and promote wound development or appear during the healing process. The importance of nutritional management has been well established in other types of chronic wounds²³ and is likely fundamental in treating leg ulcers in SCD patients. It is important to note that nephropathy or liver disease may also contribute to a low

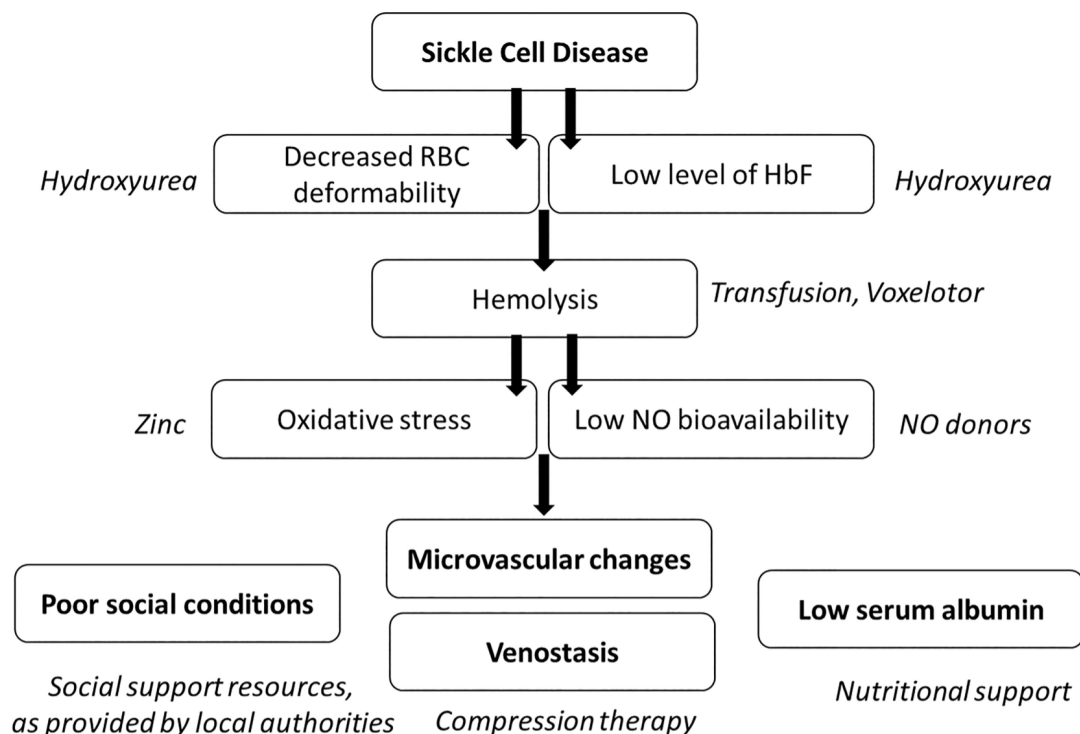


FIGURE 1 Treatments focusing on some of the mechanisms involved in leg ulcer occurrence in sickle cell disease patients.^{14,40,44,98,110,119,120}



serum albumin level.^{24,25} Therefore, laboratory tests should include measurements of albumin and pre-albumin to check for malnutrition, as well as creatinine, ALT (alanine amino transferase), ASAT (aspartate amino transferase), GGT (gamma glutamyl transferase) and alkaline phosphatase to screen for the presence of glomerulopathy and liver disease (Figure 1).

Vascular modifications

Microvascular changes can lead to inadequate tissue perfusion, an important factor in the development of chronic wounds and delayed healing. In SCD patients, microscopic examination of leg ulcer beds has revealed evidence of vasculopathy.¹⁵ This includes activated capillary endothelium, signs of thrombosis/recanalization and neovascularization around occluded capillaries^{15,26} and inflammation.¹⁵ The progression of these microvasculopathies varies depending on the patient's age, although the first ulcers may appear as early as age 10.^{5,9,12,27}

Additionally, capillary blood stasis and venostasis have been observed through microscopic examination¹⁵ and laser Doppler.²⁸ Venostasis is favoured by the abolition of the venoarteriolar reflex and cardiovascular autonomic dysfunction.²⁹ The resulting pressure from venostasis is greatest in the peri-malleolar region, which is the primary site for SCD leg ulcers.^{5,30} Stasis may also facilitate local sickling due to prolonged exposure of RBC to hypoxia and local vaso-occlusive crisis, increasing the risk of leg ulcer development.³¹ There is no evidence to support the efficacy of compression therapy with socks on leg ulcers in the absence of oedema (Figure 1). When legs are in a hanging position, inappropriate vasoconstriction has been observed around SCD ulcers,³² which could potentially contribute to delayed healing. Macrovascular abnormalities have also been observed in SCD patients with leg ulcers, suggesting involvement of diffuse vasculopathy, including increased intima media thickness in the femoral arteries.³³

The following sections will focus on the proposed mechanisms underlying vasculopathy in both the macro- and microcirculation.

Haemolysis

Sickle RBCs are prone to lysis³⁴ and the extent of haemolysis has been associated with the occurrence of leg ulcers.³⁵ Common markers of haemolysis in SCD patients with leg ulcers are high levels of LDH, bilirubin, aspartate aminotransferase and reticulocytes, concomitant with low haemoglobin levels.^{10,11,19,36,37} A recent study conducted in Senegal and Mali and that included 235 SCD adults found no difference in LDH and bilirubin levels between those with and without leg ulcers,³⁸ suggesting that haemolysis may not be the sole contributing factor to leg ulcer development, particularly in regions where clinical management and therapeutic options

differ from those in developed countries. Nevertheless, Minniti et al. observed clinical improvement in leg ulcers among SCD patients participating in the phase III, randomized, double-blinded, placebo-controlled, HOPE trial (NCT03036813), which evaluated the effects of Voxelotor.³⁹ Voxelotor is a novel molecule that inhibits HbS polymerization by increasing the affinity of HbS to oxygen (Figure 1). Voxelotor has been shown to reduce haemolysis and increase haemoglobin levels.⁴⁰ A new randomized controlled trial (RCT) called RESOLVE is currently recruiting SCD patients to specifically study the effect of Voxelotor on leg ulcer healing (NCT05561140).

Haemolysis results in reduced nitric oxide (NO) bioavailability⁴¹ and aberrant vascular activity, leading to tissue ischaemia and ulcer formation.^{15,42} In SCD patients with leg ulcers, Landburg et al. observed the elevated levels of asymmetric dimethylarginine, an inhibitor of NO synthase released during haemolysis.⁴³ Minniti et al. observed increased peri-wound cutaneous blood flow measured by laser speckle contrast imaging and infrared thermography in 18 SCD patients with leg ulcers after applying a topical NO donor.⁴⁴ Moreover, leg ulcer size and pain decreased in a dose-dependent manner (Figure 1). Ulcers healed completely in three patients who received the highest concentrations of topical sodium nitrite cream (1.8% and 2%). A phase II study is ongoing in SCD patients with leg ulcers (NCT02863068).

Reduced NO bioavailability is also associated with increased inflammation.^{45,46} Pathologists have observed an inflammatory infiltration of white blood cells in SCD ulcer tissues.^{15,26,47} The plasma levels of several pro-inflammatory cytokines (IL-1 β , IL-6, IL-8 and IL-15) were found to be significantly elevated in the serum of SCD patients with leg ulcers compared to those without.^{48–50} In an attempt to control the inflammatory process, the anti-inflammatory cytokine IL-10 level was shown to be increased in the plasma⁵¹ and in cultures of peripheral blood mononuclear cells of SCD patients with leg ulcers compared to those without.⁵²

In SCD, the most rigid RBCs are also the most fragile.³⁴ Therefore, a greater impairment in RBC deformability likely accounts for the increased haemolysis in patients with leg ulcers.^{53–55} The reduced RBC deformability could decrease tissue oxygenation and participate in leg ulcer onset.^{53–55}

Lastly, the percentage of fetal haemoglobin (HbF) determines the propensity of RBCs to sickle upon deoxygenation.⁵⁶ Thus, any factors/mechanisms that may stimulate the production of HbF may decrease haemolysis and modulate the clinical expression of SCD.⁵⁶ A number of studies have highlighted the association between low HbF levels and leg ulcer development.^{9,27,37,57,58}

HU is the most common drug used to treat SCD patients. HU improves anaemia by increasing HbF expression, which decreases HbS and its polymerization under deoxygenated conditions, thus reducing haemolysis.⁵⁹ Indeed, HU reduces RBC sickling and fragility.⁵⁹ The clinical experience with HU in patients with leg ulcers is paradoxical. While it may be expected that HU would reduce leg ulcer incidence, the literature reports several cases of leg



ulcers in patients treated with HU.^{60–63} Soya et al. reported a case of an SCD patient who developed a leg ulcer 18 days after the introduction of HU and healed after HU was stopped.⁶⁴ The authors proposed that the accumulation of HU under the skin could have caused cutaneous atrophy and impaired microcirculation.⁶⁴ An alternate hypothesis could be that the level of HbF reached during HU treatment was not enough to provide clinical benefits. Recently, Tolu et al. identified that an HbF level >25% would be necessary to limit leg ulcer development in SCD patients treated with HU,⁶⁵ a level rarely achieved. However, data from the ESCORT-HU study (a Phase IV observational cohort study including 1906 SCD patients treated with HU to the study safety profile) are reassuring. In this study, leg ulcers were reported in only 33 patients, corresponding to an incidence of 1.73%.⁶⁶ ESCORT-HU extension is currently recruiting to improve knowledge about HU and leg ulcers (NCT04707235).

Two biological/clinical phenotypes have been distinguished in SCD. One phenotype is characterized by the presence of high viscosity, which increases the risk for painful vaso-occlusive crisis, acute chest syndrome and osteonecrosis.^{35,67} The other phenotype, called the haemolytic–endothelial dysfunction phenotype, is characterized by more severe haemolysis, which leads to the development of progressive vasculopathy, and an increased risk of leg ulcers, priapism, cerebral vasculopathy, glomerulopathy and pulmonary hypertension. Many studies have shown an association between leg ulcers and echocardiographic signs of pulmonary hypertension.^{13,19,68,69} Parent et al. confirmed these results using right heart catheterization,⁷⁰ the gold standard for pulmonary hypertension diagnosis. These observations support systematic screening of pulmonary hypertension in SCD patients with leg ulcers. In addition, Nolan et al.^{36,71} and others^{10,15} have identified an association between leg ulcers and priapism.

Genetic studies

Haemoglobin genotypes, co-existing α -thalassaemia and β^S haplotypes

Powars et al. hypothesized that each SCD patient has a predetermined rate of disease progression,⁷² with genetic factors dominating the biological calendar that controls vasculopathy development (Figure 2).⁷² Four region-specific African haplotypes (Senegal, Benin, Bantu also called CAR β for Central African Republic and Cameroon haplotypes) and the Arab-India haplotype have been defined.⁷³ The CAR β haplotype has been associated with a higher incidence of soft tissue organ failure (including central nervous system, kidney, lung and leg ulcers).⁷² However, Powars et al.⁷² were the only authors to report this association, and the haplotype characterization is not systematic in SCD centres worldwide. In the Nolan study, the β^S haplotypes were not associated with leg ulcers.³⁶

Haemolytic rate also depends on haemoglobin genotype, with HbSS and HbS β^0 patients exhibiting higher level of haemolysis. Indeed, it is not surprising to observe more leg ulcers in these patients than in HbS β^+ and HbSC patients.^{5,11,27,72} Moreover, since the coexistence of α -thalassaemia modulates the degree of haemolysis, some studies logically found that patients with 2 α -gene deletions have a lower risk of developing leg ulcers.^{27,36,74} Other studies failed to find an association between leg ulcer incidence and α -thalassaemia.^{9,19,55,75} Minniti et al. proposed that these apparently conflicting results could be due to the small number of subjects with α -thalassaemia in certain studies, resulting in low statistical power, which reduces the chances of detecting a true effect.¹⁹ Thus, SCD patients without α -thalassaemia may be at an increased risk of developing leg ulcers and should be advised to consult their doctor immediately if they incur an ankle wound.

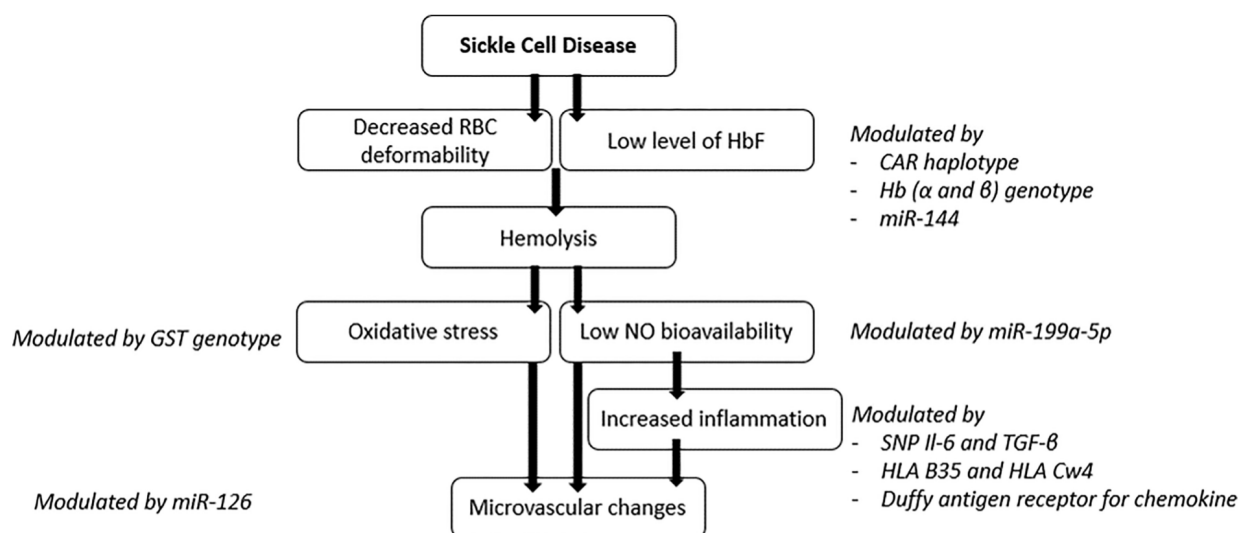


FIGURE 2 Reported genetic associations with some mechanisms involved in leg ulcer occurrence in SCD patients. GST, glutathione S transferase.



Immunity

Human leucocyte antigens (HLA) constitute a complex genetic system that encodes proteins which predominantly regulating immune/inflammatory processes. The relative risk of developing leg ulcers in SCD patients who had both HLA-B35 and HLA-Cw4 was found to be 17 times higher compared to patients without these HLAs.⁷⁶ These results suggest that genetic factors from the HLA system may contribute to the development of leg ulcers in SCD patients.⁷⁶ HLA-B35 has been linked to increased susceptibility to apoptosis⁷⁷ and endothelial dysfunction through higher production of endothelin-1 (ET-1).⁷⁸ However, the causal relationship remains elusive.

Humoral immunity may also play a role in leg ulcer risk. Namely, a single nucleotide polymorphism (SNP) of *IL6* has been proposed as a potential risk factor for leg ulcers.⁵⁰ The influence of the Duffy antigen receptor for chemokines (DARC) has also been studied in the context of SCD. DARC modulates the inflammatory process depending on its expression level. While some studies did not report any association between the occurrence of leg ulcers and DARC expression,^{79,80} Drasar et al. found an increased incidence of leg ulcers when DARC expression was high.⁸¹ It has been suggested that DARC could play a role in the recruitment of white blood cells that could participate to vascular occlusion and initiate leg ulcers.⁸¹ These findings corroborate the high level of inflammation in and around the leg ulcers mentioned above. However, there is currently no treatment targeting inflammation.

miRNA

MicroRNAs (miRNAs) are small non-coding RNAs involved in post-transcriptional gene repression by influencing messenger RNA (mRNA).⁸² The processing of primary miRNA transcripts into mature miRNAs is a regulated multistep process, and its deregulation is often associated with human diseases.⁸³

Increased expression of circulating miR-199a-5p and miR-144 has been reported in SCD patients with leg ulcers, while miR-126 expression was reduced.^{84,85} MiR-199a-5p has been shown to regulate NO production by endothelial NO-synthase in vitro⁸⁶ and may also have a regulatory role in cardiomyocytes,⁸⁷ cancer cell proliferation⁸⁸ and bile acid efflux transport.⁸⁹ MiR-144 may regulate various processes such as HbF production through gene silencing,⁸⁶ as well as cancer cell proliferation⁹⁰ and asthma.⁹¹ Lastly, miR-126, which is highly expressed in endothelial cells, plays a key role in maintaining vascular integrity and promoting angiogenesis.⁹² It is also involved in autoimmune disorders⁹³ and can be elevated by physical exercise.⁹³ Therefore, although miRNAs are considered promising biomarkers for certain pathologies, these miRNAs lack specificity and cannot be used to predict the occurrence of leg ulcers in SCD. Additionally, the regulation of miRNA involves both genetic and epigenetic factors. miRNAs interact with redox

regulation: Redox changes influence the expression of miRNAs, while miRNAs, in turn, modify the expression of redox regulators,⁹⁴ which is particularly relevant in the context of SCD. Isolating haemolysis, oxidative stress or other factors can be challenging when studying leg ulcers because these factors often interact in a complex manner, creating a 'domino' or even a 'vicious cycle' effect.

Oxidative stress

Gene regulators of oxidative stress have been studied in the context of SCD and leg ulcers. Glutathione S-transferases (GSTs) display peroxidase activity and may protect from oxidative damage. GSTs are dimeric units and, based on amino acid sequence, are described in seven classes: alpha, mu, pi, sigma, theta, omega and zeta.⁹⁵ The theta category includes GSTT1 and GSTT2 isoforms that exhibit a high activity in human RBCs. The genes encoding the Mu class (GSTM1, GSTM2, GSTM3, GSTM4 and GSTM5) appear to be active in hepatocytes and are known to be highly polymorphic. De Oliveira Filho et al. showed that patients with a GSTM1 null genotype had a 6.9-fold increased risk of leg ulcers, and patients with a GSTT1 null genotype had a 5.2-fold increased risk of leg ulcers.⁹⁶ These findings suggest that oxidative stress probably plays a role in leg ulcer pathophysiology in SCD patients. However, L-carnitine, an antioxidant, did not shorten leg ulcer healing duration in 15 SCD patients.⁹⁷ Furthermore, in a controlled trial on the effect of oral zinc sulphate in healing SCD leg ulcers, the healing rate in the treated group was three times faster than in the placebo group.⁹⁸ This study was not randomized and thus excluded from the last Cochrane systematic review on interventions for treating SCD leg ulcers.⁹⁹ However, since about one-third of SCD patients with leg ulcers have low zinc levels, Eckman proposed that all patients with active or healed ulcers should be supplemented with zinc sulphate at a dose of 220 mg administered orally three times a day.¹⁴ Zinc is an antioxidant that inhibits lipid peroxidation in RBCs, protecting the human body against oxidative stress. In addition, the oxidative stress associated with chronic haemolysis in SCD increases the demand for zinc¹⁰⁰ (Figure 1).

Furthermore, Nolan et al. studied 215 SNPs in more than 100 candidate genes selected for their potential role in SCD pathophysiology.³⁶ These included genes involved in inflammation, oxidative stress, NO biology, vasoregulation, cell-cell interaction, blood coagulation, haemostasis and growth factors. After considering a false discovery rate of 10%, only one SNP, adjacent to *SMAD7*, remained significantly associated with leg ulcers. *SMAD7* is part of the TGF- β pathway. Among its many roles, this pathway impacts cell proliferation, apoptosis, response to tissue injury, endothelial growth, inflammation, immune regulation and extracellular matrix synthesis.¹⁰¹ Although not firmly demonstrated, the TGF- β pathway could play a role in SCD leg ulcer pathophysiology. However, these results do not currently have any practical application for leg ulcer prediction and treatment.



Factors influencing healing

Genetic factors have been shown to play a role in granulation and epithelialization defects in SCD patients. Overexpression of miR-130a has been associated with chronic venous ulcers and dysregulated re-epithelization processes, granulation tissue formation and wound healing.¹⁰² MiR-130a overexpression was four times higher in SCD patients with ulcers compared to those without ulcers, suggesting miR-130a may play an important role in this complication.¹⁰³

de Carvalho-Siqueira et al. used whole exome sequencing to identify new candidate genes and found only one variant in the *FLG2* gene associated with leg ulcers in SCD patients.¹⁰⁴ This gene has been observed in persistent atopic dermatitis in African Americans and could be involved in leg ulcer development by promoting alterations in the skin barrier. At present, this finding could open a new way of understanding SCD leg ulcer pathophysiology.

At present, there is no published data using the most recent genetic approach, the genome-wide association study (GWAS). GWAS does not require any prior gene selection and allows for the study of exon and intron sequences. GWAS could be useful in identifying or confirming the involvement of genetic factors in leg ulcer mechanisms.

DELAYED HEALING OF SCD ULCERS, MAINLY VENOUS ULCERS

The second part of our review will focus on other common mechanisms shared with venous ulcers, which contribute more to healing delays than to initial ulcer occurrence (Figure 3).

In a French cohort of SCD patients, leg ulcer area < 8 cm² (odds ratio (OR) 6.73, 95% CI 2.35–19.31; *p* < 0.001) and duration < 9 weeks (OR 3.19, 95% CI 1.16–8.76; *p* = 0.024) were independently associated with wound healing at week 24 in 98 SCD patients. This study highlights that healing is more difficult in older and larger leg ulcers than in recent ones. However, it is unknown whether the natural course of these leg ulcers could have been predicted from ulcer onset or

whether the poor healing prognosis results from excessively long delay in initiating treatment. Notably, no association between wound healing and biological markers of haemolysis, HbF or other complications of SCD has been observed.¹⁰⁵

Venous involvement

Tan et al. observed haemosiderin, an iron storage complex, in the edge of the ulcers of SCD patients. The burden of haemosiderin was between the levels found in ischaemic ulcers and venous ulcers in non-SCD patients. This observation supports the idea that the pathophysiology of SCD ulcers involves both ischaemic and venous components.¹⁰⁶

Venous insufficiency

We previously highlighted the role of venostasis in the occurrence of leg ulcers in SCD patients. Venous insufficiency is also involved in delaying wound healing. Early observations indicated that bed rest promotes ulcer healing in SCD patients, suggesting that venous hyperpressure may contribute to ulcer healing delay by causing oedema.⁵ Venous hyperpressure has been evidenced by Saad and Zago, who measured the clearance (% decrease/min) of ^{99m}Tc, in SCD patients with leg ulcers to demonstrate that their calf muscle blood flow was impaired.¹⁰⁷ Additionally, Billet et al. and Mohan et al. observed rapid refill times using venous pulse recordings, venous plethysmography and laser Doppler flowmetry in SCD patients with leg ulcers compared to those without.^{32,108} To better understand the origin of venous hyperpressure, venous incompetency has been studied.

Venous incompetency

Venous hypertension is commonly associated with venous insufficiency, caused by poorly functioning incompetent venous valves. Venous incompetency can be diagnosed using Doppler duplex ultrasound to detect venous. Clare

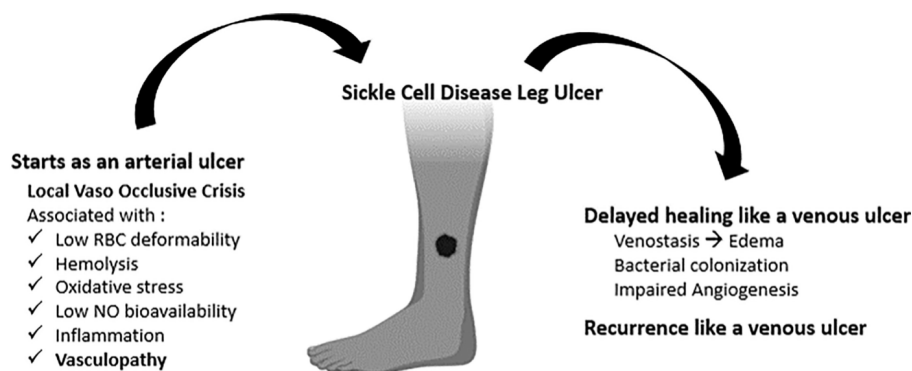


FIGURE 3 Sickle cell disease leg ulcer natural history. Sickle leg ulcer starts as arterial due to sickling, microvascular occlusion and local hypoxia. Once the ulcer is present, venostasis and impaired angiogenesis delay healing.



et al. found an increased risk of leg ulceration in SCD patients with venous incompetency, with a hazard ratio of 2.59 (95% confidence interval (CI): 2.39–2.79, $p < 0.001$).¹⁰⁹ Similarly, Cumming et al. observed a hazard ratio of 4.87 (CI: 2.09–11.03, $p < 0.001$).⁹ These data indicate that venous incompetency may contribute to the development of leg ulcers in SCD patients, but is not always present prior to ulcer onset.

Clinical application

Further investigation is required to elucidate the correlation between leg oedema and the onset of leg ulcers. Our proposed explanation is that dysfunction of the local venous and lymphatic systems render them unable to effectively manage the increased blood flow in the ulcerated area, contributing to delayed healing. The application of compression therapy is deemed essential in all SCD patients with leg ulcers, regardless of venous incompetency status.¹¹⁰ However, the question remains whether routine Doppler duplex ultrasound should be universally adopted for all SCD patients, with and without leg oedema, to screen for venous incompetency. For those SCD patients exhibiting venous incompetency, the implementation of compression therapy could serve as a preventive measure against the development of initial leg ulcers. Nevertheless, encouraging compliance in asymptomatic patients might pose a challenge.

Bacterial colonization

Bacterial colonization of leg ulcer area is an expected phenomenon that does not need to be treated by systematic or local antibiotic therapy, unless signs of local infection are present. *Staphylococcus Aureus* has been reported to be the most common bacterium present in SCD leg ulcers,^{58,111–115} as is the case in other types of ulcers.¹¹⁵ However, high microbial diversity (driven by aerobes such as *Corynebacterium* and *Alcaligenes*, and anaerobes, including *Anaerococcus*, *Peptoniphilus* and *Porphyromonas*) has been shown to delay leg ulcer healing compared to patients with less microbial diversity and a predominance of *Staphylococcus Aureus* only.¹¹⁵ On intact skin, Byeon et al. have found elevated ratios of *Corynebacterium*:*Lactobacillus* and *Staphylococcus*:*Lactobacillus* in SCD individuals with a history of leg ulcers. As no samples were collected before leg ulcer occurrence, these ratios may reflect a susceptibility to leg ulcers or consequences of healed leg ulcers. In addition, the ratio of *Lactobacillus*:*Bacillus* was elevated in SCD individuals without a history of leg ulcers. *Lactobacillus* is involved in a range of protective mechanisms against cutaneous infection or delayed healing, including immune modulation, promotion of re-epithelialization and pathogen clearance.¹¹⁶ These results, if confirmed in larger studies, could be used to target specific therapy.

Impaired angiogenesis

Nguyen et al. previously investigated wound healing differences in sickle SAD and healthy mice.¹¹⁷ They found decreased keratinocyte and fibroblast proliferation, and lower numbers of endothelial cells in SAD mice compared to healthy mice. The proliferation and mobilization of bone marrow endothelial progenitor cells were also decreased. Injecting endothelial progenitor cells into the wound shortened the healing process by promoting epidermal cell proliferation and angiogenesis. CXCL12, IL-6, VEGF A, VEGFR-2, Tie-1, Tie-2 and CXCR4 secretion was lower in the wounds of SAD mice compared to healthy mice. Interestingly, the injection of CXCL12 (a chemokine known to promote angiogenesis) also shortened the healing process.¹¹⁷ This study highlighted the well-known basal vasculopathy and impaired angiogenesis due to alterations in endothelial progenitor recruitment present in SCD. Focusing on this alteration in progenitor recruitment, autologous stem-cell therapy has been tested in 23 SCD patients with leg ulcers with promising results.¹¹⁸ However, there are no ongoing studies evaluating stem-cell therapy in a larger population.

CONCLUSION

Leg ulcers in SCD constitute a distinct category of chronic wounds with a complex and multifactorial pathophysiology that feature mechanisms commonly observed in both arterial and venous ulcers. Local vaso-occlusive crisis could trigger leg ulceration in vulnerable cutaneous areas due to underlying venostasis. Several factors may modulate the appearance of leg ulcers and healing processes, with genetic factors offering novel research opportunities to improve our understanding of the pathophysiology of this complication.

ACKNOWLEDGEMENTS

JC, NG, BF and PC wrote the paper. EN, SP and AH corrected the paper. SS revised manuscript.

CONFLICT OF INTEREST STATEMENT

All authors have no conflict of interest disclosure.

ORCID

Judith Catella  <https://orcid.org/0000-0001-8028-6827>

TWITTER

Judith Catella  judithcatella

Philippe Connes  connesphilippe

REFERENCES

1. Rees DC, Williams TN, Gladwin MT. Sickle-cell disease. *Lancet*. 2010;376(9757):2018–31.
2. Therrell BL, Lloyd-Puryear MA, Eckman JR, Mann MY. Newborn screening for sickle cell diseases in the United States: a review of data spanning 2 decades. *Semin Perinatol*. 2015;39(3):238–51.
3. Pauling L, Itano HA. Sickle cell anemia a molecular disease. *Science*. 1949;110(2865):543–8.



4. Kato GJ, Piel FB, Reid CD, Gaston MH, Ohene-Frempong K, Krishnamurti L, et al. Sick cell disease. *Nat Rev Dis Primers*. 2018;4(1):1–22.
5. Serjeant GR. Leg ulceration in sickle cell anemia. *Arch Intern Med*. 1974;133(4):690–4.
6. Alleyne SI, Wint E, Serjeant GR. Psychosocial aspects of sickle cell disease. *Health Soc Work*. 1976;1(4):104–19.
7. Alleyne SI, Wint E, Serjeant GR. Social effects of leg ulceration in sickle cell anemia. *South Med J*. 1977;70(2):213–4.
8. Powars DR, Chan LS, Hiti A, Ramicone E, Johnson C. Outcome of sickle cell anemia: a 4-decade observational study of 1056 patients. *Medicine (Baltimore)*. 2005;84(6):363–76.
9. Cumming V, King L, Fraser R, Serjeant G, Reid M. Venous incompetence, poverty and lactate dehydrogenase in Jamaica are important predictors of leg ulceration in sickle cell anaemia. *Br J Haematol*. 2008;142(1):119–25.
10. Olatunya OS, Albuquerque DM, Adekile AD, Costa FF. Evaluation of sociodemographic, clinical, and laboratory markers of sickle leg ulcers among young Nigerians at a tertiary health institution. *Niger J Clin Pract*. 2018;21(7):882–7.
11. Antwi-Boasiako C, Andemariam B, Colombatti R, Asare EV, Strunk C, Piccone CM, et al. A study of the geographic distribution and associated risk factors of leg ulcers within an international cohort of sickle cell disease patients: the CASiRe group analysis. *Ann Hematol*. 2020;99(9):2073–9.
12. Spira JAO, Borges EL, Guedes ACM, Andrade PGR, Lima VL. Prevalence of people with sickle cell disease and leg ulcers in Brazil: socioeconomic and clinical overview. *PLoS One*. 2022;17(9):e0274254.
13. Madu A, Madu K, Anigbogu I, Ugwu AO, Okwulehie VA, Ololo U, et al. Phenotypic characterisation and associations of leg ulcers in adult sickle cell patients. *Wound Repair Regen*. 2022;30(1):126–31.
14. Eckman JR. Leg ulcers in sickle cell disease. *Hematol Oncol Clin North Am*. 1996;10(6):1333–44.
15. Minniti CP, Delaney KMH, Gorbach AM, Xu D, Lee CCR, Malik N, et al. Vasculopathy, inflammation, and blood flow in leg ulcers of patients with sickle cell anemia. *Am J Hematol*. 2014;89(1):1–6.
16. Durosinmi MA, Gevao SM, Esan GJ. Chronic leg ulcers in sickle cell disease: experience in Ibadan. *Nigeria Afr J Med Sci*. 1991;20(1):11–4.
17. Delaney KMH, Axelrod KC, Buscetta A, Hassell KL, Adams-Graves PE, Seamon C, et al. Leg ulcers in sickle cell disease: current pattern and practices. *Hemoglobin*. 2013;37(4):325–32.
18. Reis de Souza V, Kelly S, Cerdeira Sabino E, Mendes de Oliveira F, Silva T, Miranda Teixeira C, et al. Factors associated with leg ulcers in adults with sickle cell disease in Brazil. *Adv Skin Wound Care*. 2023;36(2):98–105.
19. Minniti CP, Taylor JG, Hildesheim M, O'Neal P, Wilson J, Castro O, et al. Laboratory and echocardiography markers in sickle cell patients with leg ulcers. *Am J Hematol*. 2011;86(8):705–8.
20. Shi C, Dumville JC, Cullum N, Connaughton E, Norman G. Compression bandages or stockings versus no compression for treating venous leg ulcers. *Cochrane Database Syst Rev*. 2021;7(7):CD013397.
21. Zhang H, Yang Z. Research on dynamic comfort maintenance by measuring lower limb oedema and seat pressure during simulated seated sleep in flight. *Int J Occup Saf Ergon*. 2024;30(1):72–83.
22. Molnar JA, Underdown MJ, Clark WA. Nutrition and chronic wounds. *Adv Wound Care (New Rochelle)*. 2014;3(11):663–81.
23. Stechmiller JK. Understanding the role of nutrition and wound healing. *Nutr Clin Pract*. 2010;25(1):61–8.
24. Ataga KL, Saraf SL, Derebail VK. The nephropathy of sickle cell trait and sickle cell disease. *Nat Rev Nephrol*. 2022;18(6):361–77.
25. Kato GJ, Steinberg MH, Gladwin MT. Intravascular hemolysis and the pathophysiology of sickle cell disease. *J Clin Invest*. 2017;127(3):750–60.
26. Serjeant GR, Serjeant BE, Mohan JS, Clare A. Leg ulceration in sickle cell disease: medieval medicine in a modern world. *Hematol Oncol Clin North Am*. 2005;19(5):943–56.
27. Koshy M, Entsuaeh R, Koranda A, Kraus AP, Johnson R, Bellvue R, et al. Leg ulcers in patients with sickle cell disease. *Blood*. 1989;74(4):1403–8.
28. Gniadecka M, Gniadecki R, Serup J, Søndergaard J. Microvascular reactions to postural changes in patients with sickle cell anaemia. *Acta Derm Venereol*. 1994;74(3):191–3.
29. Oguanobi N, Onwubere B, Anisiuba B, Ike S, Ejim E, Ibegbulam O. Clinical findings associated with cardiovascular autonomic dysfunction in adult sickle cell anaemia patients. *Acta Cardiol*. 2012;67:169–75.
30. Hassan A, Gayus GL, Abdulrasheed I, Umar MA, Ismail DL, Babadoko AA. Chronic leg ulcers in sickle cell disease patients in Zaria. *Nigeria Arch Int Surg*. 2014;4(3):141–5.
31. Gabuzda TG. Sickle cell leg ulcers: current pathophysiologic concepts. *Int J Dermatol*. 1975;14(5):322–5.
32. Mohan JS, Marshall JM, Reid HL, Thomas PW, Serjeant GR. Postural vasoconstriction and leg ulceration in homozygous sickle cell disease. *Clin Sci (Lond)*. 1997;92(2):153–8.
33. Ayoola OO, Bolarinwa RA, Onakpoya UU, Adedeji TA, Onwuka CC, Idowu BM. Intima-media thickness of the common femoral artery as a marker of leg ulceration in sickle cell disease patients. *Blood Adv*. 2018;2(22):3112–7.
34. Connes P, Lamarre Y, Waltz X, Ballas SK, Lemonne N, Etienne-Julan M, et al. Haemolysis and abnormal haemorheology in sickle cell anaemia. *Br J Haematol*. 2014;165(4):564–72.
35. Kato GJ, Gladwin MT, Steinberg MH. Deconstructing sickle cell disease: reappraisal of the role of hemolysis in the development of clinical subphenotypes. *Blood Rev*. 2007;21(1):37–47.
36. Nolan VG, Adewoye A, Baldwin C, Wang L, Ma Q, Wyszynski DF, et al. Sickle cell leg ulcers: associations with Haemolysis and SNPs in klotho, TEK and genes of the TGF- β /BMP pathway. *Br J Haematol*. 2006;133(5):570–8.
37. Bartolucci P, Brugnara C, Teixeira-Pinto A, Pissard S, Moradkhani K, Jouault H, et al. Erythrocyte density in sickle cell syndromes is associated with specific clinical manifestations and hemolysis. *Blood*. 2012;120(15):3136–41.
38. Ranque B, Diaw M, Dembele AK, Lapoumeroulie C, Offredo L, Tessougue O, et al. Association of haemolysis markers, blood viscosity and microcirculation function with organ damage in sickle cell disease in sub-Saharan Africa (the BIOCADRE study). *Br J Haematol*. 2023;203(2):319–26.
39. Minniti CP, Knight-Madden J, Tonda M, Gray S, Lehrer-Graiwer J, Biemond BJ. The impact of voxelotor treatment on leg ulcers in patients with sickle cell disease. *Am J Hematol*. 2021;96(4):E126–E128.
40. Vissa M, Vichinsky E. Voxelotor for the treatment of sickle cell disease. *Expert Rev Hematol*. 2021;14(3):253–62.
41. Chirico EN, Pialoux V. Role of oxidative stress in the pathogenesis of sickle cell disease. *IUBMB Life*. 2012;64(1):72–80.
42. Trent JT, Kirsner RS. Leg ulcers in sickle cell disease. *Adv Skin Wound Care*. 2004;17(8):146–410.
43. Landburg PP, Teerlink T, Biemond BJ, Brandjes DPM, Muskiet FAJ, Duits AJ, et al. Plasma asymmetric dimethylarginine concentrations in sickle cell disease are related to the hemolytic phenotype. *Blood Cell Mol Dis*. 2010;44(4):229–32.
44. Minniti CP, Gorbach AM, Xu D, Hon YY, Delaney KM, Seidel M, et al. Topical sodium nitrite for chronic leg ulcers in patients with sickle cell anaemia: a phase 1 dose-finding safety and tolerability trial. *Lancet Haematol*. 2014;1:e95–e103.
45. Riddell DR, Owen JS. Nitric oxide and platelet aggregation. *Vitam Horm*. 1997;57:25–48.
46. De Caterina R, Libby P, Peng HB, Thannickal VJ, Rajavashisth TB, Gimbrone MA, et al. Nitric oxide decreases cytokine-induced endothelial activation. Nitric oxide selectively reduces endothelial expression of adhesion molecules and proinflammatory cytokines. *J Clin Invest*. 1995;96(1):60–8.



47. Francillon YJ, Jilly PN, Varricchio F, Castro O. Histochemical analysis of growth factor, fibronectin, and iron content of sickle cell leg ulcers. *Wound Repair Regen.* 1996;4(2):240–3.
48. Bowers AS, Reid HL, Greenidge A, Landis C, Reid M. Blood viscosity and the expression of inflammatory and adhesion markers in homozygous sickle cell disease subjects with chronic leg ulcers. *PLoS One.* 2013;8(7):e68929.
49. Domingos IF, Pereira-Martins DA, Sobreira MJVC, Oliveira RTD, Alagbe AE, Lanaro C, et al. High levels of proinflammatory cytokines IL-6 and IL-8 are associated with a poor clinical outcome in sickle cell anemia. *Ann Hematol.* 2020;99(5):947–53.
50. Vicari P, Adegoke SA, Mazzotti DR, Caçado RD, Nogutti MAE, Figueiredo MS. Interleukin-1 β and interleukin-6 gene polymorphisms are associated with manifestations of sickle cell anemia. *Blood Cell Mol Dis.* 2015;54(3):244–9.
51. Belisário AR, Mendes-Oliveira F, de Souza VR, Bolina-Santos E, Mendes FG, Moreno EC, et al. Association between inflammatory molecules, nitric oxide metabolites and leg ulcers in individuals with sickle cell anemia. *Hematol Transfus Cell Ther.* 2022;44(2):169–76.
52. Rêgo MJBM, da Silva RR, Pereira MC, da Silva Araújo A, da Rocha Pitta MG, Falcão DA, et al. Evaluation of CD4⁺CD25⁺FoxP3⁺ T cell populations, IL-10 production, and their correlation with clinical and biochemical parameters in sickle cell anemia patients with leg ulcers. *Cytokine.* 2015;75(2):310–5.
53. Ballas SK. Sickle cell anemia with few painful crises is characterized by decreased red cell deformability and increased number of dense cells. *Am J Hematol.* 1991;36(2):122–30.
54. Bowers AS, Duncan WW, Pepple DJ. Red blood cell deformability is reduced in homozygous sickle cell disease patients with leg ulcers. *Clin Hemorheol Microcirc.* 2016;5(2):199–204.
55. Connes P, Lamarre Y, Hardy-Dessources MD, Lemonne N, Waltz X, Mouguel D, et al. Decreased hematocrit-to-viscosity ratio and increased lactate dehydrogenase level in patients with sickle cell anemia and recurrent leg ulcers. *PLoS One.* 2013;8(11):e79680.
56. Akinshey I, Alsultan A, Solovieff N, Ngo D, Baldwin CT, Sebastiani P, et al. Fetal hemoglobin in sickle cell anemia. *Blood.* 2011;118(1):19–27.
57. Hutz MH, Salzano FM, Adams J, Opitz JM. Hb F levels, longevity of homozygotes and clinical course of sickle cell anemia in Brazil. *Am J Med Genet.* 1983;14(4):669–76.
58. Adedeji MO, Ukoli FA. Haematological factors associated with leg ulcer in sickle cell disease. *Trop Geogr Med.* 1987;39(4):354–6.
59. Charache S, Terrin ML, Moore RD, Dover GJ, Barton FB, Eckert SV, et al. Effect of hydroxyurea on the frequency of painful crises in sickle cell anemia. Investigators of the multicenter study of hydroxyurea in sickle cell anemia. *N Engl J Med.* 1995;332(20):1317–22.
60. Dissemmond J, Hoefft D, Knab J, Franckson T, Kroger K, Goos M. Leg ulcer in a patient associated with hydroxyurea therapy. *Int J Dermatol.* 2006;45(2):158–60.
61. Hwang SW, Hong SK, Kim SH, Seo JK, Lee D, Sung HS. A hydroxyurea-induced leg ulcer. *Ann Dermatol.* 2009;21(1):39–41.
62. Best PJ, Daoud MS, Pittelkow MR, Pettitt RM. Hydroxyurea-induced leg ulceration in 14 patients. *Ann Intern Med.* 1998;128(1):29–32.
63. Salmon-Ehr V, Leborgne G, Vilque JP, Potron G, Bernard P. Effets secondaires cutanés de l'hydroxyurée: étude prospective de 26 patients consultant dans un service de dermatologie. *Rev Med Interne.* 2000;21(1):30–4.
64. Soya E, Makowski C, Blaise S. Leg ulcer induced by hydroxycarbamide in sickle cell disease: what is the therapeutic impact? *Int Wound J.* 2019;16(4):897–902.
65. Tolu SS, Crouch A, Choi J, Gao Q, Reyes-Gil M, Ogu UO, et al. Hydroxyurea and fetal hemoglobin effect on leg ulcers in patients with sickle cell disease. *Ann Hematol.* 2022;101(3):541–8.
66. de Montalembert M, Voskaridou E, Oevermann L, Cannas G, Habibi A, Loko G, et al. Real-life experience with hydroxyurea in patients with sickle cell disease: results from the prospective ESCORT-HU cohort study. *Am J Hematol.* 2021;96(10):1223–31.
67. Alexander N, Higgs D, Dover G, Serjeant GR. Are there clinical phenotypes of homozygous sickle cell disease? *Br J Haematol.* 2004;126(4):606–11.
68. Serarslan G, Akgül F, Babayigit C. High prevalence of pulmonary hypertension in homozygous sickle cell patients with leg ulceration. *Clin Exp Hypertens.* 2009;31(1):44–8.
69. De Castro LM, Jonassaint JC, Graham FL, Ashley-Koch A, Telen MJ. Pulmonary hypertension associated with sickle cell disease: clinical and laboratory endpoints and disease outcomes. *Am J Hematol.* 2008;83(1):19–25.
70. Parent F, Bachir D, Inamo J, Lionnet F, Driss F, Loko G, et al. A hemodynamic study of pulmonary hypertension in sickle cell disease. *N Engl J Med.* 2011;365(1):44–53.
71. Nolan VG, Baldwin C, Ma Q, Wyszynski DF, Amirault Y, Farell JJ, et al. Association of single nucleotide polymorphisms in *kltho* with priapism in sickle cell anaemia. *Br J Haematol.* 2004;128:266–72.
72. Powars D, Chan LS, Schroeder WA. The variable expression of sickle cell disease is genetically determined. *Semin Hematol.* 1990;27(4):360–76.
73. Flint J, Harding RM, Boyce AJ, Clegg JB. The population genetics of the haemoglobinopathies. *Baillieres Clin Haematol.* 1998;11(1):1–51.
74. Higgs DR, Aldridge BE, Lamb J, Clegg JB, Weatherall DJ, Hayes RJ, et al. The interaction of alpha-thalassaemia and homozygous sickle-cell disease. *N Engl J Med.* 1982;306(24):1441–6.
75. Steinberg MH. Predicting clinical severity in sickle cell anaemia. *Br J Haematol.* 2005;129(4):465–81.
76. Ofofu MD, Castro O, Alarif L. Sickle cell leg ulcers are associated with HLA-B35 and Cw4. *Arch Dermatol.* 1987;123(4):482–4.
77. Salazar G, Colombo G, Lenna S, Antonioli R, Beretta L, Santaniello A, et al. HLA-B35 influences the apoptosis rate in human peripheral blood mononucleated cells and HLA-transfected cells. *Hum Immunol.* 2007;68(3):181–91.
78. Santaniello A, Salazar G, Lenna S, Antonioli R, Colombo G, Beretta L, et al. HLA-B35 upregulates the production of endothelin-1 in HLA-transfected cells: a possible pathogenetic role in pulmonary hypertension. *Tissue Antigens.* 2006;68(3):239–44.
79. Nebor D, Durpes MC, Mouguel D, Mukisi-Mukaza M, Elion J, Hardy-Dessources MD, et al. Association between Duffy antigen receptor for chemokines expression and levels of inflammation markers in sickle cell anemia patients. *Clin Immunol.* 2010;136(1):116–22.
80. Afenyi-Annan A, Kail M, Combs MR, Orringer EP, Ashley-Koch A, Telen MJ. Lack of Duffy antigen expression is associated with organ damage in patients with sickle cell disease. *Transfusion.* 2008;48(5):917–24.
81. Drasar ER, Menzel S, Fulford T, Thein SL. The effect of Duffy antigen receptor for chemokines on severity in sickle cell disease. *Haematologica.* 2013;98(8):e87–e89.
82. Lee RC, Feinbaum RL, Ambros V. The *C. Elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell.* 1993;75(5):843–54.
83. Kim VN, Han J, Siomi MC. Biogenesis of small RNAs in animals. *Nat Rev Mol Cell Biol.* 2009;10(2):126–39.
84. Santos ED, Melo GI, Santana PV, Quadros IG, Yahuouédéhou SC, Guarda CC, et al. A description of the hemolytic component in sickle leg ulcer: the role of circulating miR-199a-5p, miR-144, and miR-126. *Biomol Ther.* 2022;12(2):317.
85. Le NT, Abe JI. MicroRNA 199a and the eNOS (endothelial NO synthase)/NO pathway. *Arterioscler Thromb Vasc Biol.* 2018;38(10):2278–80.
86. Li B, Zhu X, Ward CM, Starlard-Davenport A, Takezaki M, Berry A, et al. MIR-144-mediated NRF2 gene silencing inhibits fetal hemoglobin expression in sickle cell disease. *Exp Hematol.* 2019;70:85–96.
87. Zhang H, Li S, Zhou Q, Sun Q, Shen S, Zhou Y, et al. Qiliqiangxin attenuates phenylephrine-induced cardiac hypertrophy through downregulation of MiR-199a-5p. *Cell Physiol Biochem.* 2016;38(5):1743–51.
88. Hua Q, Jin M, Mi B, Xu F, Li T, Zhao L, et al. LINC01123, a c-Myc-activated long non-coding RNA, promotes proliferation and aerobic



- glycolysis of non-small cell lung cancer through miR-199a-5p/c-Myc axis. *J Hematol Oncol.* 2019;12(1):91.
89. Balasubramanian N, Devereaux M, Orlicky D, Sokol R, Suchy F. miR-199a-5p inhibits the expression of ABCB11 in obstructive cholestasis. *J Biol Chem.* 2021;297(6):101400. <https://pubmed.ncbi.nlm.nih.gov/34774795/>
 90. Kooshkaki O, Rezaei Z, Rahmati M, Vahedi P, Derakhshani A, Brunetti O, et al. MiR-144: a new possible therapeutic target and diagnostic/prognostic tool in cancers. *Int J Mol Sci.* 2020;21(7):2578.
 91. Rodrigo-Muñoz JM, Gil-Martínez M, Lorente-Sorolla C, García-Latorre R, Valverde-Monge M, Quirce S, et al. miR-144-3p is a biomarker related to severe corticosteroid-dependent asthma. *Front Immunol.* 2022;13:858722.
 92. Jansen F, Yang X, Hoelscher M, Cattelan A, Schmitz T, Proebsting S, et al. Endothelial microparticle-mediated transfer of microRNA-126 promotes vascular endothelial cell repair via SPRED1 and is abrogated in glucose-damaged endothelial microparticles. *Circulation.* 2013;128(18):2026–38.
 93. Ma Y, Liu H, Wang Y, Xuan J, Gao X, Ding H, et al. Roles of physical exercise-induced MiR-126 in cardiovascular health of type 2 diabetes. *Diabetol Metab Syndr.* 2022;4(1):169.
 94. Leisegang MS, Schröder K, Brandes RP. Redox regulation and non-coding RNAs. *Antioxid Redox Signal.* 2018;29(9):793–812.
 95. Silva DG, Junior EB, de Souza Torres L, Júnior OR, de Castro Lobo C, Bonini-Domingos CR, et al. Relationship between oxidative stress, glutathione S-transferase polymorphisms and hydroxyurea treatment in sickle cell anemia. *Blood Cell Mol Dis.* 2011;47(1):23–8.
 96. de Oliveira Filho RA, Silva GJ, de Farias Domingos I, Domingues Hatzlhofer BL, da Silva AA, de Lima Filho JL, et al. Association between the genetic polymorphisms of glutathione S-transferase (GSTM1 and GSTT1) and the clinical manifestations in sickle cell anemia|Elsevier enhanced reader. *Blood Cell Mol Dis.* 2013;51:76–9.
 97. Serjeant BE, Harris J, Thomas P, Serjeant GR. Propionyl-L-carnitine in chronic leg ulcers of homozygous sickle cell disease: a pilot study. *J Am Acad Dermatol.* 1997;37(3 Pt 1):491–3.
 98. Serjeant GR, Galloway RE, Gueri MC. Oral zinc sulphate in sickle-cell ulcers. *Lancet.* 1970;2(7679):891–2.
 99. Martí-Carvajal AJ, Knight-Madden JM, Martínez-Zapata MJ. Interventions for treating leg ulcers in people with sickle cell disease. *Cochrane Database Syst Rev.* 2021;1(1):CD008394.
 100. Miranda CT, Vermeulen-Serpa KM, Pedro AC, Brandão-Neto J, de Lima Vale SH, Figueiredo MS. Zinc in sickle cell disease: a narrative review. *J Trace Elem Med Biol.* 2022;72:126980.
 101. Attisano L, Wrana JL. Signal transduction by the TGF-beta superfamily. *Science.* 2002;296(5573):1646–7.
 102. Pastar I, Khan AA, Stojadinovic O, Lebrun EA, Medina MC, Brem H, et al. Induction of specific MicroRNAs inhibits cutaneous wound healing. *J Biol Chem.* 2012;287(35):29324–35.
 103. Batista THC, Santana RM, Sobreira MJ, Arcanjo GS, Domingos IF, Pereira-Martins DA, et al. Up-regulation of miR-130a is related to leg ulcers in sickle cell anaemia. *Br J Haematol.* 2022;197(1):e16–e18.
 104. de Carvalho-Siqueira GQ, Ananina G, de Souza BB, Borges MG, Ito MT, da Silva-Costa SM, et al. Highlight article: whole-exome sequencing indicates FLG2 variant associated with leg ulcers in Brazilian sickle cell anemia patients. *Exp Biol Med (Maywood).* 2019;244(11):932–9.
 105. Senet P, Blas-Chatelain C, Levy P, Manea EM, Peschanski M, Mirault T, et al. Factors predictive of leg-ulcer healing in sickle cell disease: a multicentre, prospective cohort study. *Br J Dermatol.* 2017;177(1):206–11.
 106. Tan J, Smith A, Abisi S, Eastham D, Burnand KG. Tissue and urinary haemosiderin in chronic leg ulcers. *Eur J Vasc Endovasc Surg.* 2007;34(3):355–60.
 107. Saad STO, Zago MA. Leg ulceration and abnormalities of calf blood flow in sickle-cell anemia. *Eur J Haematol.* 1991;46(3):188–90.
 108. Billet HH, Patel Y, Rivers SP. Venous insufficiency is not the cause of leg ulcers in sickle cell disease. *Am J Hematol.* 1991;37(2):133–4.
 109. Clare A, FitzHenley M, Harris J, Hambleton I, Serjeant GR. Chronic leg ulceration in homozygous sickle cell disease: the role of venous incompetence. *Br J Haematol.* 2002;119(2):567–71.
 110. Minniti CP, Kato GJ. Critical reviews: how we treat sickle cell patients with leg ulcers. *Am J Hematol.* 2016;91(1):22–30.
 111. Babalola OA, Ogunkeyede A, Odetunde AB, Fasola F, Oni AA, Babalola CP, et al. Haematological indices of sickle cell patients with chronic leg ulcers on compression therapy. *Afr J Lab Med.* 2020;9(1):8.
 112. MacFarlane DE, Baum KF, Serjeant GR. Bacteriology of sickle cell leg ulcers. *Trans R Soc Trop Med Hyg.* 1986;80(4):553–6.
 113. Ademiluyi SA, Rotimi VO, Coker AO, Banjo TO, Akinyanju O. The anaerobic and aerobic bacterial flora of leg ulcers in patients with sickle-cell disease. *J Infect.* 1988;17(2):115–20.
 114. Sehgal SC, Arunkumar BK. Microbial flora and its significance in pathology of sickle cell disease leg ulcers. *Infection.* 1992;20(2):86–8.
 115. Byeon J, Blizinsky KD, Persaud A, Findley K, Lee JJ, Buscetta AJ, et al. Insights into the skin microbiome of sickle cell disease leg ulcers. *Wound Repair Regen.* 2021;29(5):801–9.
 116. Mohammedsaed W, Cruickshank S, McBain AJ, O'Neill CA. Lactobacillus rhamnosus GG lysate increases Re-epithelialization of keratinocyte scratch assays by promoting migration. *Sci Rep.* 2015;5:16147.
 117. Nguyen VT, Nassar D, Batteux F, Raymond K, Tharaux PL, Aractingi S. Delayed healing of sickle cell ulcers is due to impaired angiogenesis and CXCL12 secretion in skin wounds. *J Invest Dermatol.* 2016;136(2):497–506.
 118. Meneses JVL, Fortuna V, de Souza ES, Daltro GC, Meyer R, Minniti CP, et al. Autologous stem cell-based therapy for sickle cell leg ulcer: a pilot study. *Br J Haematol.* 2016;175(5):949–55.
 119. Lemonne N, Charlot K, Waltz X, Ballas SK, Lamarre Y, Lee K, et al. Hydroxyurea treatment does not increase blood viscosity and improves red blood cell rheology in sickle cell anemia. *Haematologica.* 2015;100(10):e383–e386.
 120. Pule GD, Mowla S, Novitzky N, Wiysonge CS, Wonkam A. A systematic review of known mechanisms of hydroxyurea-induced fetal hemoglobin for treatment of sickle cell disease. *Expert Rev Hematol.* 2015;8(5):669–79.

How to cite this article: Catella J, Guillot N, Nader E, Skinner S, Poutrel S, Hot A, et al. Controversies in the pathophysiology of leg ulcers in sickle cell disease. *Br J Haematol.* 2024;00:1–10. <https://doi.org/10.1111/bjh.19584>

